



WEBINAR SERIES



BREEDERS TALK GREEN

GENETIC DIVERSITY AND RESPONSIBLE ANIMAL BREEDING

Thursday 1 July 2021 | 13:30 - 15:00 CET



The role of Aquaculture Breeders in Maintaining Genetic Diversity



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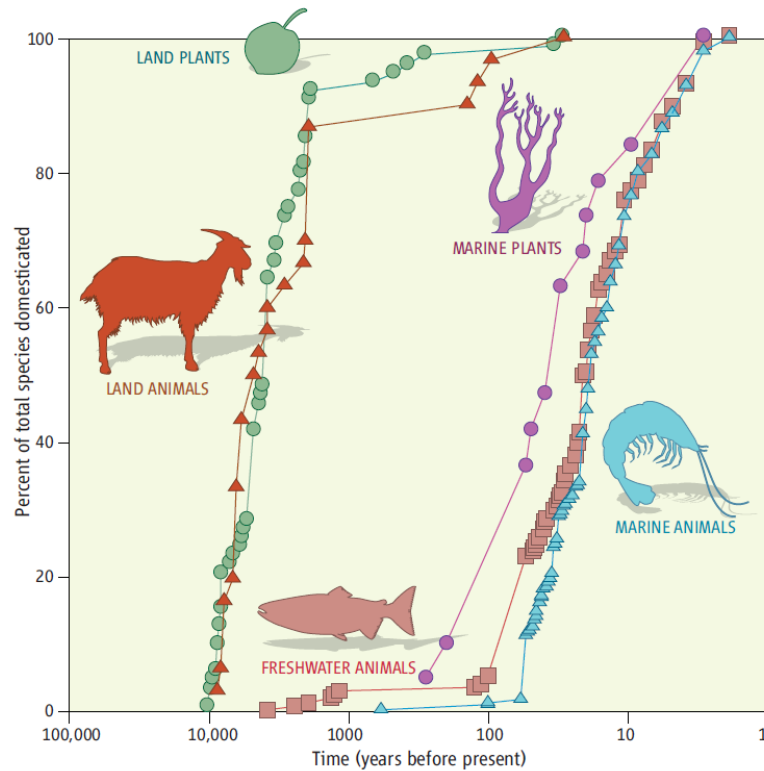


SYSAAF (French poultry and aquaculture breeding association)

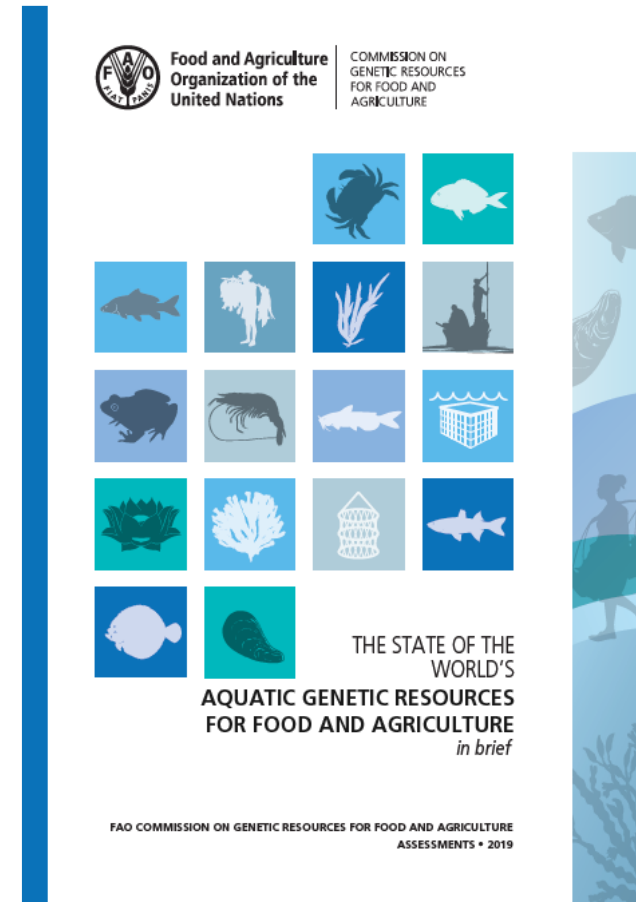
- Non profit organisation (French law on farming)
- 27 geneticists and data scientists (4 PhD students)
- 31 breeding companies (18 aquaculture)
- 30 species (11 fishes, 4 molluscs, 4 shrimps)
- Genetic indexation of 119 genetic lines
- International expertise : Thailand, Mozambique, Mexico, Madagascar



Aquaculture has just initiated domestication by the creation of genetic resources



Duarte et al., 2007



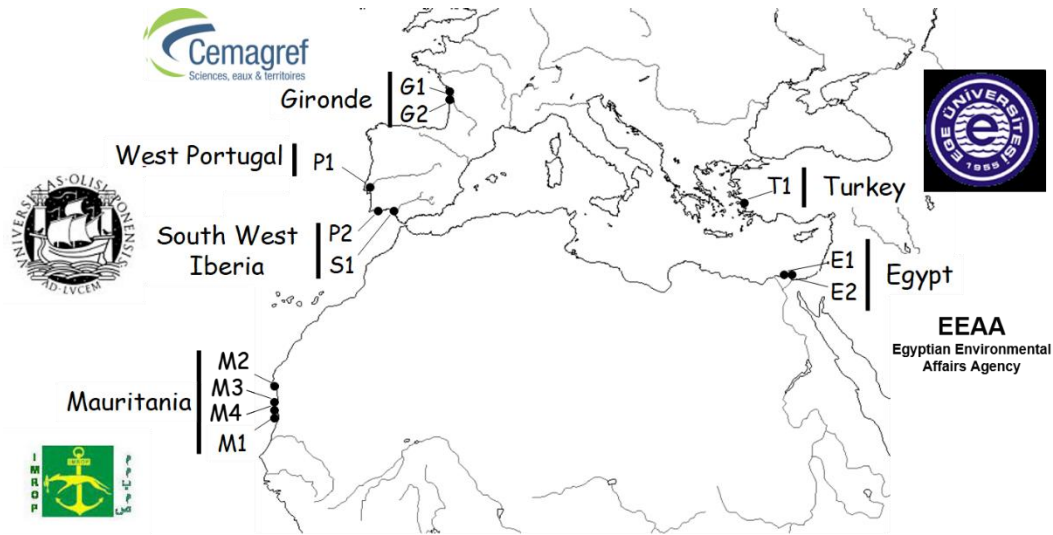
<http://www.fao.org/3/ca5256en/CA5256EN.pdf>

Core message is that aquaculture breeders are actively involved in the management of genetic diversity

- In characterising genetic variability of their stocks
- In using genomic tools to trace their pedigrees (family relationship) in using DNA fingerprinting
- In adapting software to improve pedigree assignment with DNA fingerprinting
- In applying genetic principles to optimise conservation of genetic diversity
- In cryopreserving their genetic resources for long term private and public conservation

Evaluation of genetic variability for successful domestication

- Domestication of the meagre *Argyrosomus regius*
- Collection of tissues samples for genotyping (n=100)

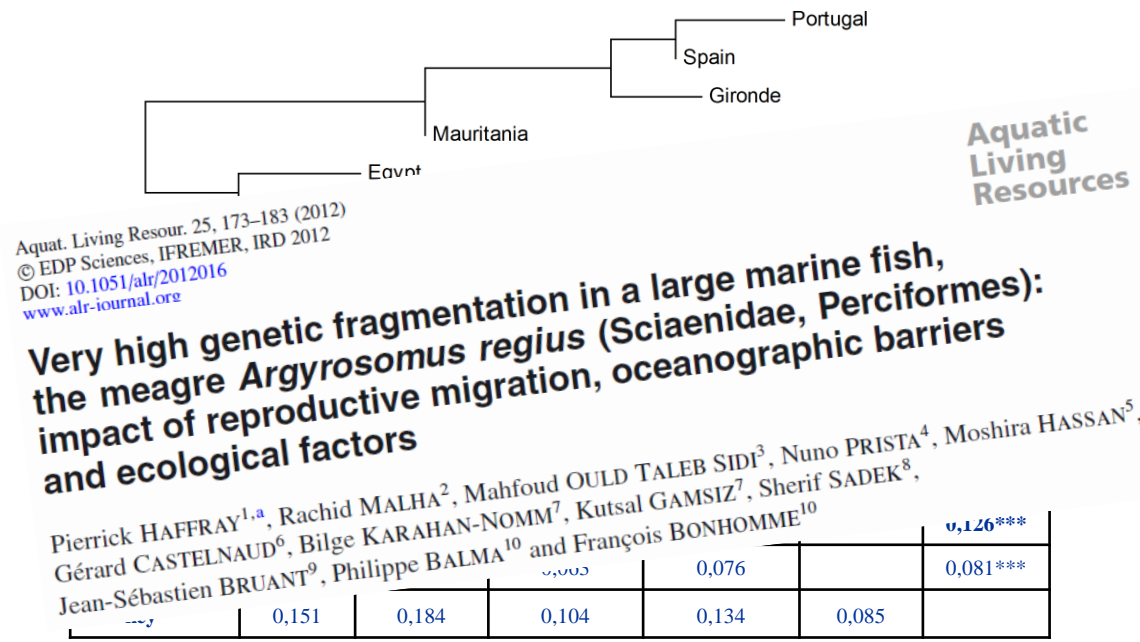


- ABS (Access and Benefit Sharing) agreement with EEAA (Egypt)

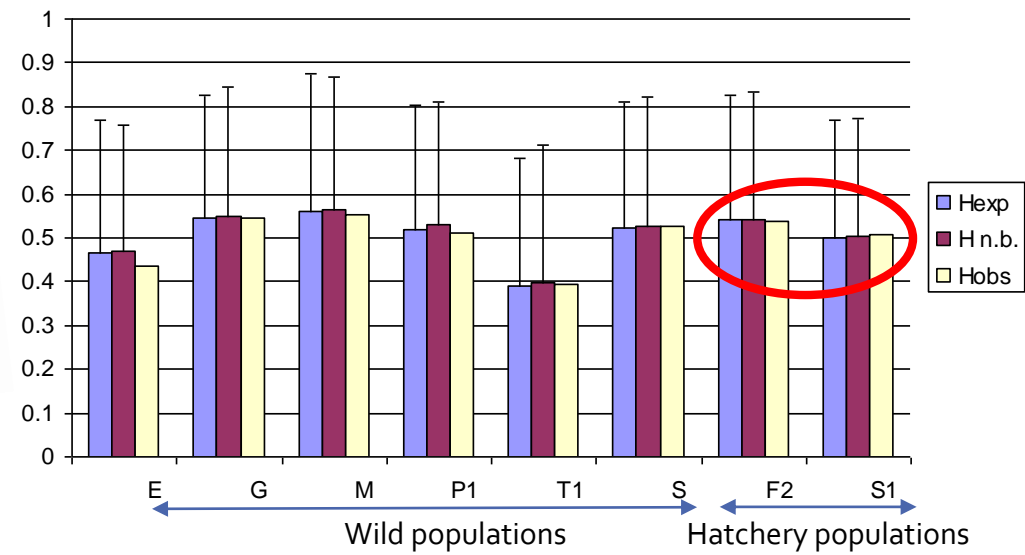


Majors results

- Very important genetic differentiation between wild populations (interspecific ?)



Haffray et al., 2012



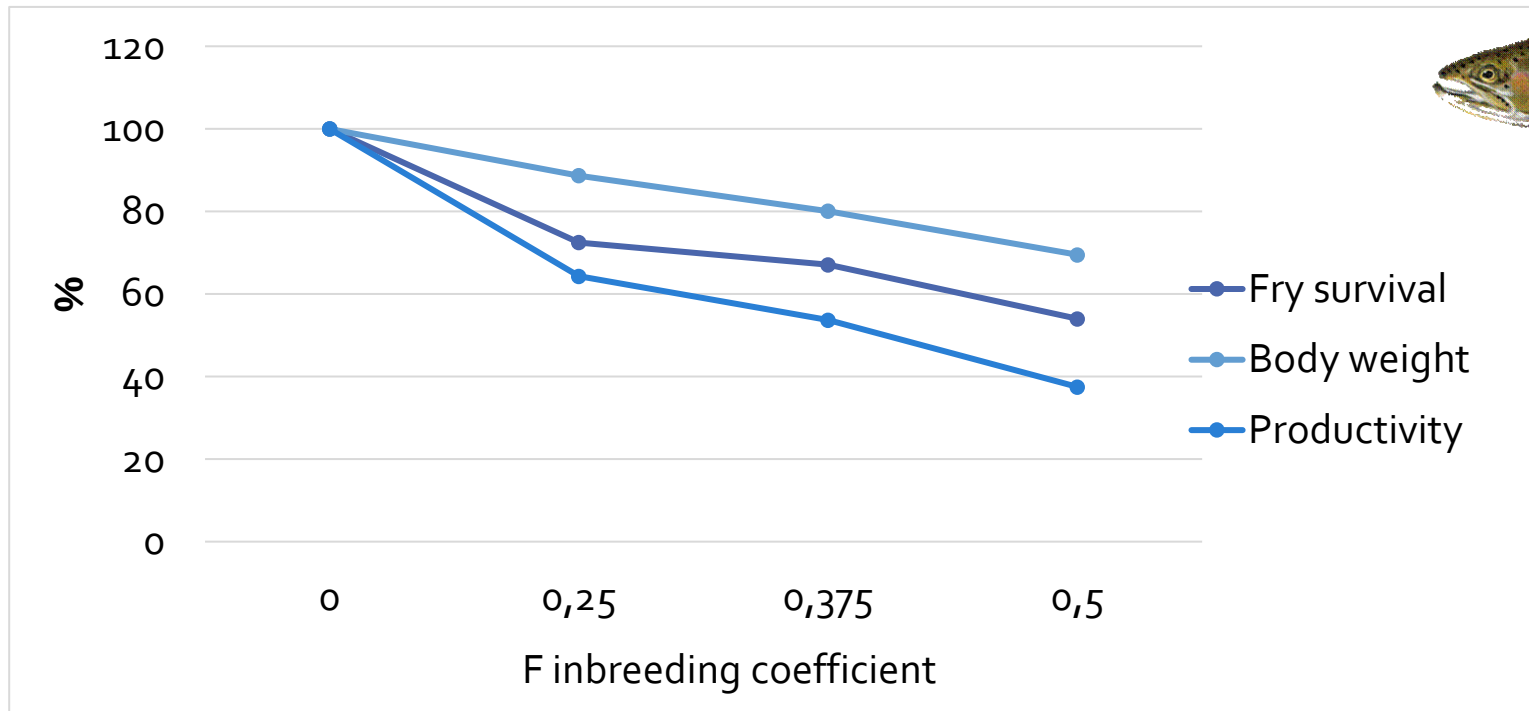
Haffray et al., 2013 (EAS, Las Palmas)

- French aquacultural Fo stocks have similar levels of genetic variability than wild populations



Risks associated with uncontrolled inbreeding?

- Evaluation by repeated crosses between brothers and sisters during 3 generations



Gjerde et al., 1983
Rainbow trout
3 successive generation of brother-sister mating

- Additionnal negative effects (malformation, sensibility to diseases, adaptation)
- There is no long term sustainable breeding programs without managing inbreeding

General principles of a breeding program in aquaculture

1. Creation of the 200/3200 families / generation

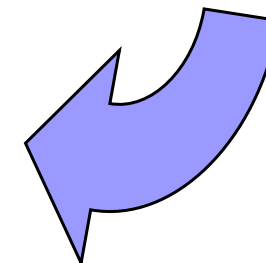
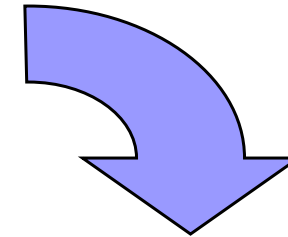
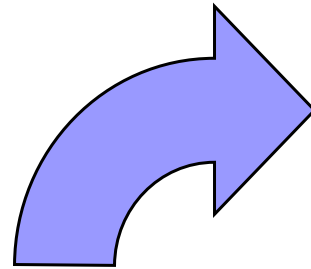
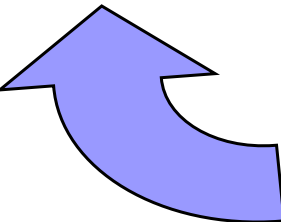


2. Separate family rearing until individual tagging with electronic transponder and pooling in 1 tank (cage)

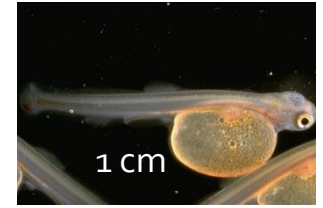
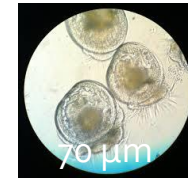


3. Rearing and phenotyping of candidates and sibs
disease resistance, processing, flesh quality, reproductive traits...

4. Genetic and genomic indexation and proposal of best candidates maximising genetic progress and minimizing inbreeding



How to get pedigree in aquaculture?



“Traditional” Initial separated family rearing



- Individual electronic tagging (50 to 1000 / family) before family mixing
- Initial investment in facilities, personal
- Potential high tank effect

“Innovative” Family pooled at hatching



French favorite approach as more adapted to SMEs

- DNA collection, individual tagging
- DNA fingerprinting (7-8 € / individual)
- No specific facilities but genotyping cost
- No tank effect = more precise

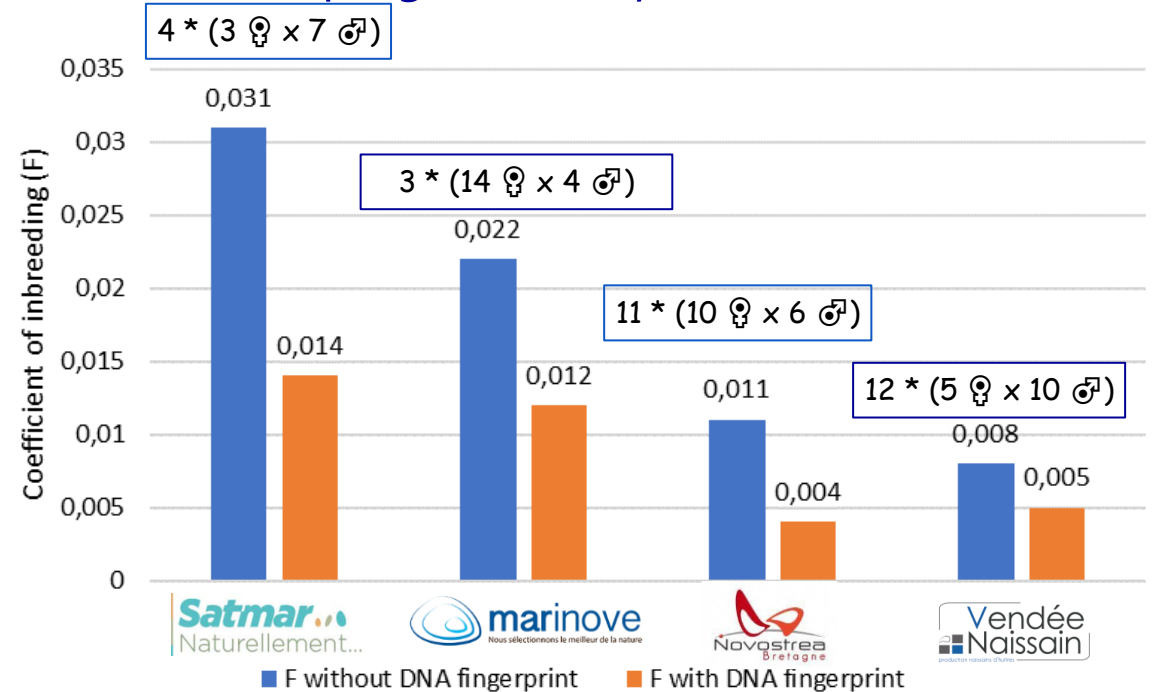


Introduction of DNA fingerprinting in shellfish breeding



- Development of a panel to assign pedigree in Pacific oyster *Crassostrea gigas*
- Evaluation of breeding practices in commercial selection programs in 1 year class

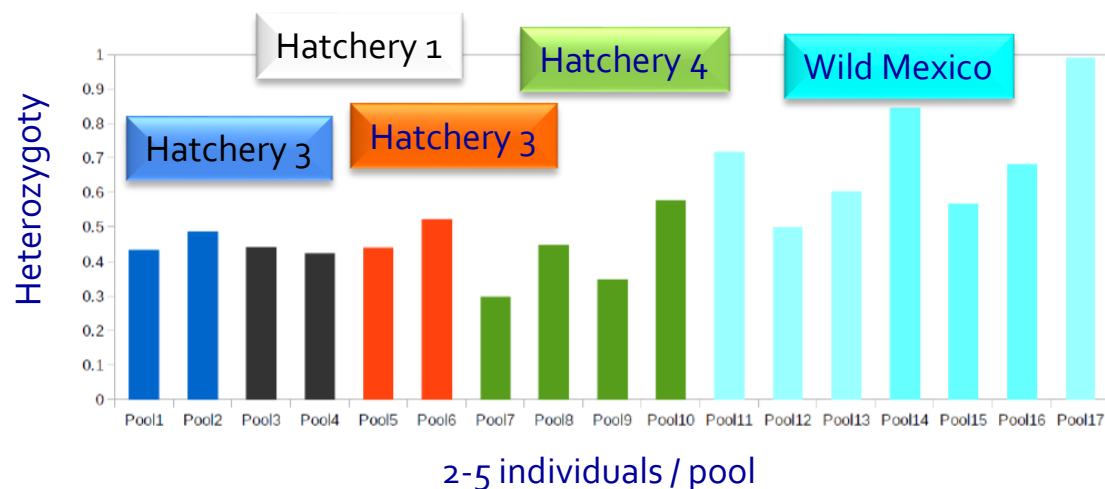
MOLECULAR ECOLOGY RESOURCES
 Molecular Ecology Resources (2014) 14, 820–830
Development of SNP-genotyping arrays in two shellfish species
 doi: 10.1111/1755-0998.1223
 S. LAPÈGUE,* E. HARRANG,* S. HEURTEBISE,* E. FLAHAUW,* C. DONNADIEU,† P. GAYRAL,‡§
 †* RAILLENGHIEN,‡ L. GENESTOUT,¶ L. BARBOTTE,¶ R. MAHLA,¶ P. HAFFRAY** and C. KLOPP*



- Introduction of DNA fingerprinting is helping to secure domestication in managing inbreeding risk

Introduction of DNA fingerprinting in shrimp breeding

- The blue shrimp *Litopenaeus stylirostris* was introduced in New Caledonia (1700 T/year) in the 70' with a limited effective of 3-5 founders (Goyard et al., 2003)
- Failure to introduce new genetic resources from sources not resistant to IHHN virus
- Automation of 171 SNP markers from 459 669 locus RAD identified by RAD sequencing



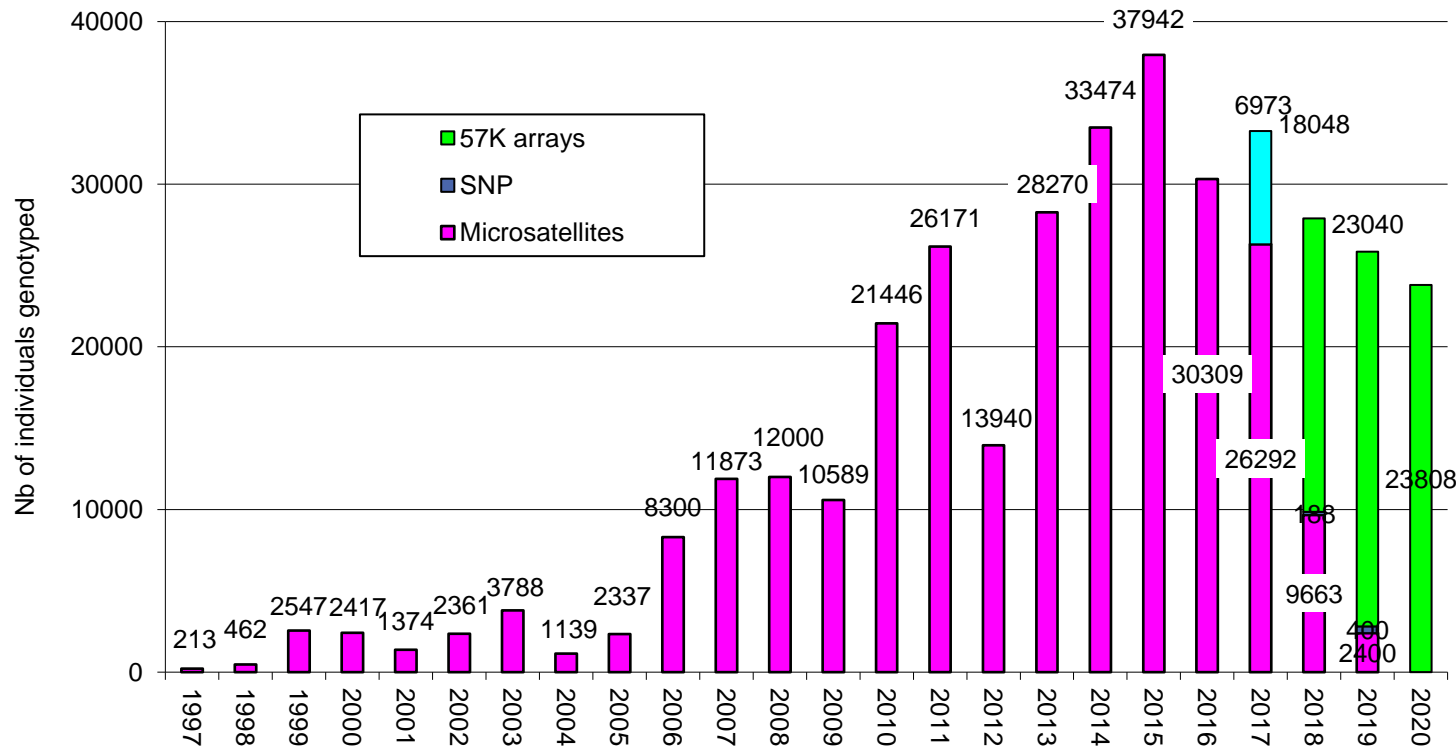
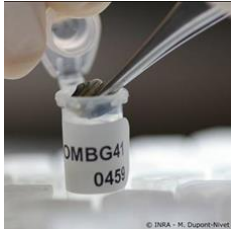
Trait	Raw shrimp		Cooked shrimp	
	Mean (Std)	Heritability (S.E.)	Mean (Std)	Heritability (S.E.)
Weight (g)	14.5 (3.0)	0.52 (0.11)	14.0 (3.1)	0.52 (0.11)
Length (mm)	129.6 (7.9)	0.46 (0.10)	-	-
L*	48.1 (2.4)	0.41 (0.10)	72.1 (1.9)	0.45 (0.10)
a*	1.2 (0.8)	0.59 (0.11)	23.3 (2.7)	0.53 (0.11)
b*	6.4 (3.4)	0.51 (0.10)	19.1 (2.7)	0.54 (0.11)
Tail yield	-	-	0.59 (0.03)	0.22 (0.06)

1200 progenies DNA assigned from 37 ♀ x 31 ♂

Enez et al., 2018 (WCGALP)

- The limited genetic variability(-30-50 % / wild) is not preventing future genetic progress

DNA parentage assignment is now widely applied in SYSAAF to improve breeding programs



- 368 692 fishes, shrimp, or molluscs genotyped since 1997

- 8 microsatellite panels for parentage assignment (trout, sea bass, sea bream, turbot, meagre, red drum, salmon, siberian sturgeon)
- 8 SNP panels for parentage assignment (meagre, blue shrimp, tiger shrimp, white shrimp, Pacific oyster, Japanese clam, rainbow trout)
- 5 HD 50 K ThermoFisher and Illumina arrays (trout, sea bass, sea bream, Pacific oyster, white shrimp)

- 1 QTL VNN

Adaptation of software to assign parentage in an octoploid (8N) species, the Siberian sturgeon *Acipenser baeri* for caviar production

- The French caviar production (40 T) is based on two introductions from Russia (1975, 1982) with potentially limited genetic variability

- Adaptation of the VITASSIGN software

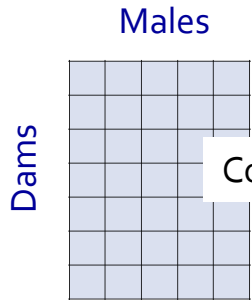
- allowing
- with



ploidy level :

ants

2 alleles for (1) to be true



Genetic parameters of caviar yield, color, size and firmness using parentage assignment in an octoploid fish species, the Siberian sturgeon *Acipenser baerii*

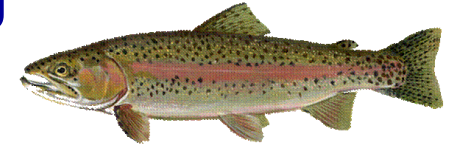
A. Bestin^{a,*}, O. Brunel^b, A. Malledant^b, B. Debeuf^b, P. Benoit^b, R. Mahla^c, H. Chapuis^{f,g}, D. Guémené^{f,h}, M. Vandeputte^{d,e}, P. Haffray^a

Body weight	Oocyte size	Caviar yield	Caviar colour
0.07	0.37 ± 0.06	0.52 ± 0.04	0.49 ± 0.08
0.06	0.36 ± 0.06	0.66 ± 0.07	0.42 ± 0.06

Heritabilities (Bestin et al., 2021)
(496 females genotyped with microsatellite markers)

- It is possible to DNA assign in commercial conditions an 8N animal species to improve stocks and manage inbreeding

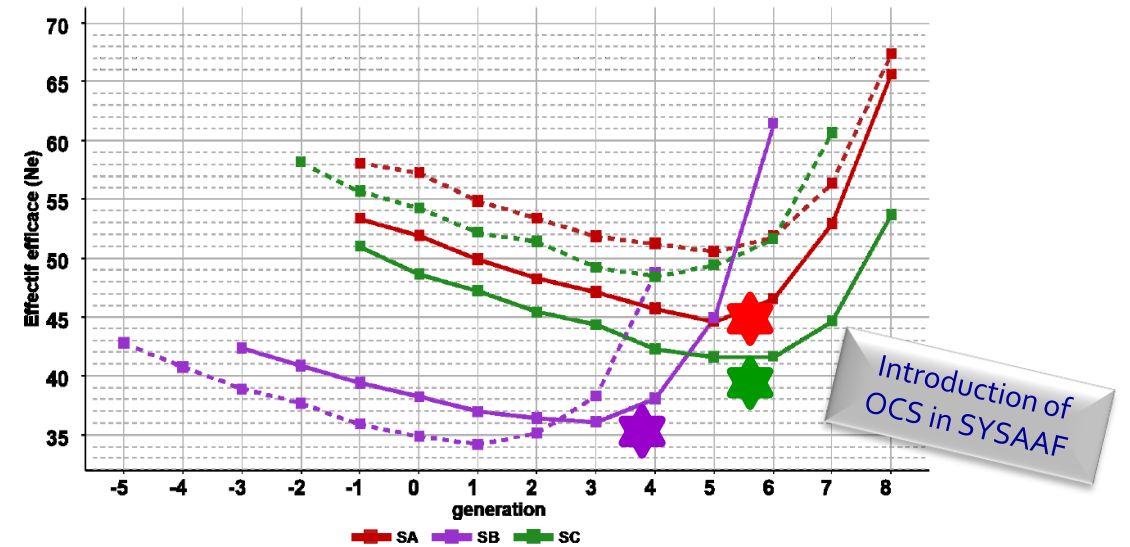
Optimisation of mating to minimise increase of inbreeding



- How to choose to mate who with who to create the new generation?
- Application of the principle of “Optimal contribution selection” (OCS) adapted to poultry and aquaculture (Chapuis et al., 2016) in SYSAAF since 2012 (adaptative simulated annealing - ASA)
- Genotyping of the 2 last generations from 3 breeding programs on 57K SNP array



- Evaluation of the Linkage Disequilibrium of long homozygous conserved DNA fragments (ROH)

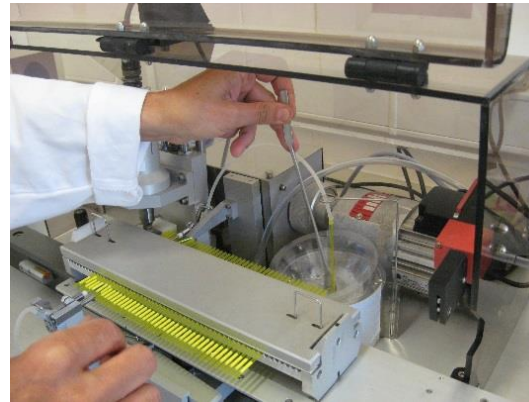


D'Ambrosio et al., (2019) and D'Ambrosio (2020)

The use of adapted OCS-ASA improve the conservation of the genetic variability

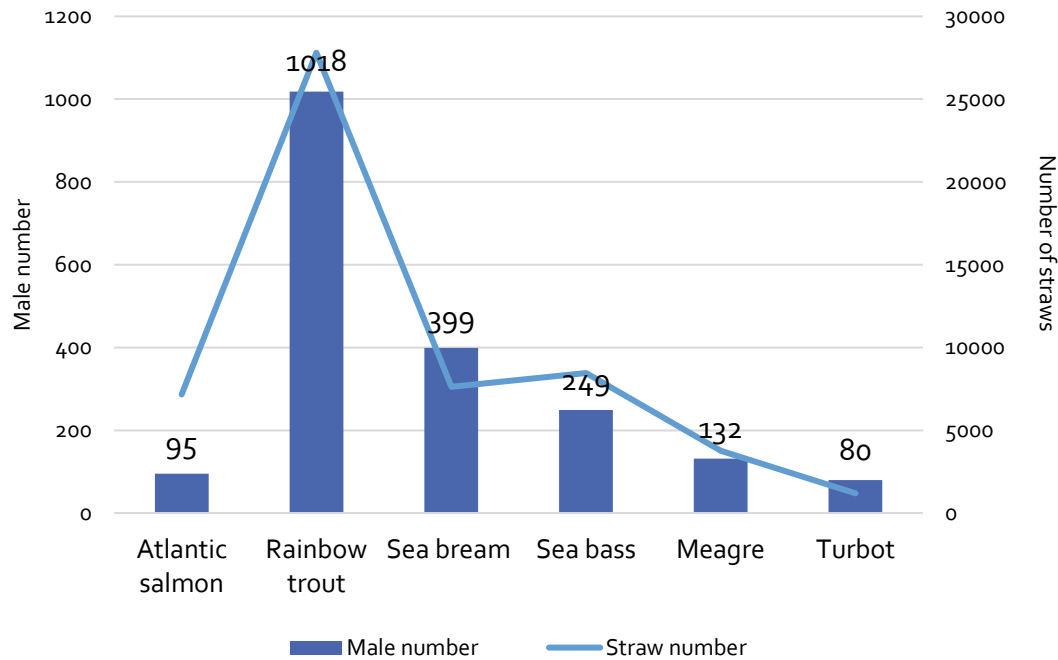
Long term conservation of genetic in using cryopreservation

- Initial R&D works to developed extenders and established freezing and conservation procedures (Haffray et al., 2008)
- Partnership to develop a collective cryobank cryoaqua operational since 2012



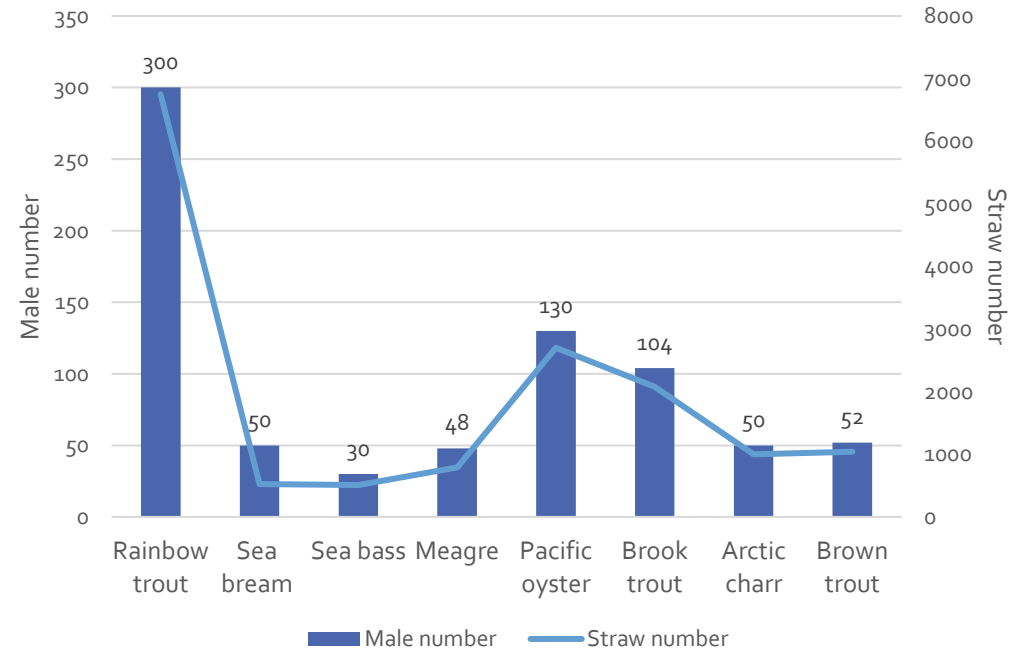
Long term conservation of genetic in using cryopreservation

Private cryobanking at CRYOAQUA

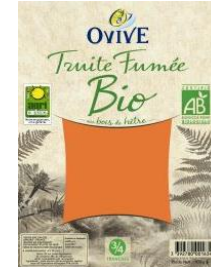


Public cryobanking at

CRYOBANQUE NATIONALE
Groupement d'Intérêt Scientifique



- Sperm cryopreservation is routinely operated to conserve genetic resources in France
- Partnership with Cryobanque Nationale is securing long term conservation of genetic resources



Thank you for your attention

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